

# Rachel K Osborn

CURRICULIM VITAE (February 2021)

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## EDUCATION

**PhD: Entomology** 2014-present  
Michigan State University, East Lansing, MI  
Research: *Diversity of South American ambrosia beetles (Curculionidae: Scolytinae: Xyleborini) and their fungal partners*  
Candidacy achieved September 2018  
Advisor: Anthony Cognato

**Bachelor of Arts: Conservation Biology, Entomology** 2003-2008  
Hampshire College, Amherst, MA  
Thesis: *The evolution of insect-fungus symbiosis*

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## INSECT COLLECTIONS EXPERIENCE

**Graduate Research Assistant**, Holistic Insect Systematics Laboratory, Michigan State University 2016-2019  
Contributed to digitization efforts of the MSU AJ Cook Arthropod Research Collection  
Transferred specimen records to the Symbiota Collections of Arthropod Network ([SCAN](#))  
Archived ~1000 specimen records from ethanol-preserved larvae, slides, and pinned specimens  
Supervisors: *Gary Parsons, Anthony Cognato*

**Graduate Research Assistant**, Holistic Insect Systematics Laboratory, Michigan State University 2016-2018  
Assisted on US Farm Bill project producing a [key](#) to Southeast Asian Xyleborini (Curculionidae: Scolytinae)  
Maintained and improved scientific specimen photography equipment  
Designed and implemented workflow to generate montage imaged of insect specimens  
Produced >1200 photographs of xyleborine beetles for use in online interactive identification key  
Supervisors: *Sarah Smith, Anthony Cognato*

**Graduate Research Assistant**, Holistic Insect Systematics Laboratory, Michigan State University 2014-2016  
Collaborated on the Advancing Digitization of Biodiversity Collections Project  
Organized data and trained other in use of a Linux-based whole-drawer imaging robot  
Created digital photographs of whole drawers of insects from the MSU AJ Cook Arthropod Research Collection  
Supervisors: *Gary Parsons, Anthony Cognato*

**Photographer of Entomology types**, Smithsonian National Museum of Natural History, Washington, DC 2011-2014  
Cooperated with curators across entomology departments: Hymenoptera, Coleoptera, Diptera, and Hemiptera  
Generated montaged digital images of holotype specimens for the [USNM EMu](#) and [Hymtypes](#) databases  
Supervisors: *David Furth, Patricia Gentili-Poole, Michael Gates, Robert Kula, Matthew Buffington, Steven Lingafelter, Christian Thompson, Torsten Dikow, Thomas Henry*

**Entomology Type Database Contributor**, Smithsonian National Museum of Natural History 2007-2011  
Researched and vetted type information for the USNM Lepidoptera and Hymenoptera holotype collections  
Compiled type description citations and collection records for integration into the museum's EMu catalog  
Supervisors: *John Brown, Alma Solis, Michael Gates, Robert Kula, Matthew Buffington*

**INSECT COLLECTIONS EXPERIENCE (Continued)**

**Seasonal Employee**, Maryland Department of Natural Resources 2008-2009  
 Located moths from Maryland in the Smithsonian USNM entomological collection  
 Assembled moth collection data into a Microsoft Access database  
 Supervisor: *Jennifer Frye*

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**COLLECTIONS AND INSECT TAXONOMY WORKSHOPS**

**Natural Science Collections Fellowship** (\$6,000), Michigan State University 2019  
 Participated in cross-disciplinary discussions about Natural History collection management and maintenance  
 Curated the Mecoptera in the MSU AJ Cook Arthropod Research Collection  
 Constructed a digital and physical teaching collection of local insects identified to species  
 Instructors: *Barbara Lundrigan, L. Alan Prather, Anthony Cognato*

**Entomological Collections Management Workshop**, Milwaukee Public Museum, Milwaukee, WI 2017  
 Took part in interactive workshop sponsored by the Entomological Collections Network  
 Joined in discussions and hand-on training focused on the curation and managements of insect collections  
 Instructors/Coordinators: *Floyd Shockley, David Furth, Jennifer Zaspel*

**Ecology and Evolution of Coleoptera (Beetles)**, La Selva Biological Station, Costa Rica 2017  
 Studied taxonomy and identification of Coleoptera in course organized by the Organization for Tropical Studies  
 Discovered field collection techniques for tropical ecosystems  
 Assessed the diversity of beetles associated with fungal fruiting bodies at La Selva, Costa Rica  
 Fabricated, deployed and serviced flight intercept traps for bark beetles at the biological station  
 Instructors: *Richard Leschen, Chris Carlton, Nathan Lord, Victoria M. Bayless*

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**RESEARCH EXPERIENCE**

**Sesiidae Phylogeny Project**, Holistic Insect Systematics Laboratory, Michigan State University 2018-present  
 Supported laboratory research efforts  
 Documented project progress and organized lab mores and records  
 Extracted, amplified and sequenced DNA from 97 clearwing moths  
 Collaborators: *Anthony Cognato, William Taft*

**Early Detection and Rapid Response Project**, Michigan State University 2014-2017  
 Sorted bark and ambrosia beetles from baited trap residue  
 Assigned preliminary identification to collected bark and ambrosia beetles  
 Supervisor: *Anthony Cognato*

**Molecular Forest Ecology**, Hulcr Laboratory, School of Forest Resources and Conservation, University of Florida 2015  
 Developed aptitude for collecting ambrosia beetles and isolating their fungi  
 Retrieved GenBank sequence data from ambrosia fungi for analysis  
 Hosts: *Jiri Hulcr, Adam Black, Caroline Storer*

**NSF Research Experience for Undergraduates Research Intern**, Harvard Forest, Petersham, MA 2006  
 Curated the Formicidae collection at the Harvard Forest Station  
 Measured and analyzed soil respiration data at the Harvard Forest LTER Site  
 Conducted biodiversity survey of Coleoptera and Formicidae at the Harvard Forest LTER Site  
 Supervisor: *Aaron Ellison*

**RESEARCH EXPERIENCE (Continued)**

**NSF Research Experience for Undergraduates Research Intern**, Mountain Lake Biological Station, Pembroke, VA 2005  
 Observed and recorded the nocturnal mating and dispersal behavior of *Bolitotherus cornutus* (Tenebrionidae)  
 Supervisor: *Patrice Ludwig*

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**RESEARCH TRAVEL**

**Pontificia Universidad Católica del Ecuador (PUCE)**, Quito Ecuador July 28 – August 4, 2019  
 Extracted and amplified DNA from cultured fungal samples

**Pontificia Universidad Católica del Ecuador (PUCE)**, Quito Ecuador January 14 – March 18, 2018  
 Initiated collaborative relationships with collections staff at the university  
 Liaised with Biological Stations staff and governmental employees to organize field research  
 Hired and supervised indigenous community member to provide research assistance in the field  
 Collected ambrosia beetles in the provinces of Orellana and Napo  
 Obtained collection and mobilization permits for research materials  
 Isolated ambrosia fungi from collected beetles at PUCE in Quito

**Universidad Técnica Estatal de Quevedo (UTEQ)**, Quevedo, Ecuador March 1 – 28, 2017  
 Gained practical knowledge about molecular fungal culturing techniques to isolate ambrosia fungi  
 Collected bark and ambrosia beetles in the provinces of Los Rios, Cotopaxi, and Guayas, Ecuador  
 Isolated ambrosia fungi from collected beetles at UTEQ laboratory in Quevedo  
 Informed UTEQ students about the diversity and ecological importance of bark and ambrosia beetles  
 Assisted in training government employees in scolytine identification and population monitoring

**Universidad Técnica Estatal de Quevedo (UTEQ)**, Quevedo, Ecuador May 10 – 23, 2015  
 Assisted in formation of cooperative relationship between UTEQ and MSU for Cognato Lab research in Ecuador  
 Collected bark and ambrosia beetles in the provinces of Los Rios, and Cotopaxi, Ecuador  
 Isolated ambrosia fungi from collected beetles at UTEQ laboratory in Quevedo  
 Trained UTEQ students how to collect bark and ambrosia beetles

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**TEACHING EXPERIENCE**

**Teaching Assistant:** MSU ISB 201L *Insects, Globalization and Sustainability Laboratory* SP2020, FA2019  
 Taught biology to non-science major undergraduates SP2016, FA 2015  
 Added content and polished lectures  
 Created quizzes and exams  
 Performed general classroom maintenance including cleaning, setup and breakdowns of hands-on activities  
 Instructed 12 sections of 27 students each  
 Supervisors: Gabe Ording, Ryan Kimbirauskas, Amanda Lorenz

**Curriculum Development:** *Diversity and taxonomy of insects*

Developed classroom unit for 875 non-science major undergraduate students  
 Fabricated lecture introducing systematics and taxonomy, basic insect classification, anatomy, and identification  
 Constructed peer-led classroom game to familiarize students with the defining characters of major insect orders  
 Designed cooperative activity to teach students how to identify major insect orders using modified taxonomic key

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**GRANTS AND AWARDS (total \$20,705)**

Graduate School Writing Fellowship in the Disciplines, Michigan State University; (\$2,000)	2019-2020
Research Enhancement Award, Michigan State University College of Graduate Studies; (\$2,005)	2019
Natural Science Collections Fellowship, Michigan State University; (\$6,000)	2019
Robert R. Dreisbach Endowed Memorial Fellowship, Dept. of Entomology, Michigan State University; (\$2,000)	2019
J.E. and Jean M. McPherson Graduate Student Award, Dept. of Entomology, Michigan State University; (\$1,200)	2018
Bug House Fellowship, Dept. of Entomology, Michigan State University; (\$500)	2017
Hutson Endowment for PhD Award, Dept. of Entomology, Michigan State University; (\$2,500)	2016
President's Prize, First place, student poster competition – phylogeny; Entomological Society of America Systematics, Evolution and Biodiversity	2015
Graduate Student Research Enrichment Award, Coleopterists Society; (\$3,000)	2014
Award for Graduate Student Research, Society of Systematic Biologists; (\$1,500)	2014

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**PUBLICATIONS**

Osborn, R. 2020. [At MSU a 153-year-old insect collection is being used to solve modern problems.](#) *Lansing State Journal*. August 7, 2020.

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**ORAL PRESENTATIONS**

Osborn, R., and A.I. Cognato. 2019. Relations across kingdoms: Ecuadorian *Coptoborus* and *Theoborus* (Curculionidae: Scolytinae: Xyleborini) and their fungal partners. SOGA: Sacred Order of the Genuiculate Antennae. Annual meeting of the Entomological Society of America, St. Louis, MO, USA

Osborn, R., M. Jusino, C. Bateman, and A.I. Cognato. 2018. Continuing illumination of the South America ambrosia beetle (Coleoptera: Curculionidae: Scolytinae) diversity and their fungal associates. Graduate student oral competition. Annual meeting of the Entomological Society of America, Vancouver, BC, Canada.

Osborn, R. 2017. Diversidad de escarabajos de ambrosias xyleborina sudamericanos y sus hongos asociados. Universidad Técnica Estatal de Quevedo. Quevedo, Ecuador.

Osborn, R. 2016. Preliminary phylogeny of the South American xyleborine genera *Coptoborus* and *Theoborus* (Coleoptera: Curculionidae: Scolytinae). Graduate student oral competition. International Congress of Entomology, Orlando, FL, USA

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**POSTER PRESENTATIONS**

Osborn, R., and A.I. Cognato. 2015. Preliminary phylogeny of the South American xyleborine genera *Coptoborus* and *Theoborus* (Coleoptera: Curculionidae: Scolytinae), poster. Entomological Society of America Meeting, Minneapolis, MN, USA

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**MENTORSHIP**

Peer mentees, Cognato Lab: *Erica Fischer*, Masters Student (2018-2020); *Gina Sari*, Masters Student, (2016-2019)  
 Estudiantes de ingeniería, Universidad Técnica Estatal de Quevedo: *Joselyn J Bas*, (2015-2017); *Denny Carriel*, (2015-2017); *Eduardo Gutierrez Lara*; (2017); *Jonathan Chavez* (2017); *Shirley Thyago* (2017)

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## OUTREACH AND SERVICE

- Student Group for Diversity, Equity and Inclusion**, Dept. of Entomology, Michigan State University 2020-present  
 Read books and articles regarding social and cultural inclusion in academia and scientific research  
 Publicized group meetings to students in the department and encouraged participation  
 Supported and participated in thoughtful group discussions  
 Organized meeting logistics and choice of discussion topics
- Skype a Scientist** 2020-present  
 Explained my research on ambrosia beetles to elementary school-aged students via video chat  
 Engaged students' questions about ecology, evolutionary biology, and entomology
- Science Writing for News Outlets**, Graduate School, Michigan State University 2020  
 Trained in basic journalism skills including interviewing, ethics, and science writing for the public  
 Developed a focused reporting plan for news article advocating for the importance of natural history collections  
 Published news story for general audience about the function of the MSU insect collection in research  
 Mentor: *Matt Miller*, Editor and Storytelling Coach for the Lansing State Journal and Battle Creek Observer
- Bug House Fellow**, Dept. of Entomology, Michigan State University 2014-2018  
 Presented to visitors about insect anatomy and biology  
 Instructed visitors how to hold and interact with live specimens  
 Trained less experienced guides how to give Bug House tours
- Curriculum Committee**, Dept. of Entomology, Michigan State University 2015-2016  
 Contributed to departmental initiatives to standardize course syllabi and student learning outcomes
- Partners in Evolution volunteer**, Smithsonian National Museum of Natural History, Washington, DC 2008-2009  
 Educated visitors to live butterfly exhibit about butterfly biology
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## LANGUAGES

- English – Native fluency  
 Spanish – Basic working proficiency
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## PROFESSIONAL MEMBERSHIPS

- Lepidopterists' Society**, *since February 2019*  
**Society for the Preservation of Natural History Collections**, *since July 2017*  
**Entomological Collections Network**, *since November 2016*  
**Entomological Society of America**, *since November 2014*  
**Coleopterists' Society**, *since January 2015*
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